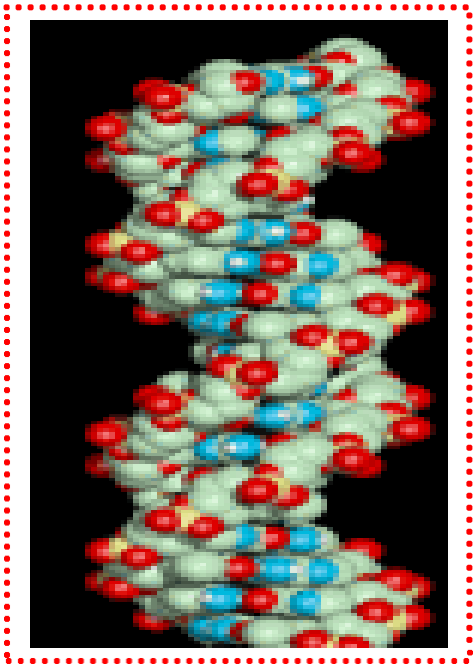
UNIVERSITY OF DELAWARE

COLLEGE OF HEALTH SCIENCES

DEPARTMENT OF MEDICAL AND MOLECULAR SCIENCES



PROGRAM IN APPLIED MOLECULAR BIOLOGY AND BIOTECHNOLOGY

PRACTICUM HANDBOOK

2024-25 *08/2024*

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SECTION 1 – ADMINSTRATIVE INFORMATION

UNIVERSITY OF DELAWARE
COLLEGE OF HEALTH SCIENCES
DEPARTMENT OF MEDICAL AND MOLECULAR SCIENCES
PROGRAM IN APPLIED MOLECULAR BIOLOGY AND BIOTECHNOLOGY

COURSE NUMBERS AND TITLES:

(Senior students enrolled in the Baccalaureate Program register for these courses for the academic terms indicated)

MMSC 441 Biotechnology Practicum I:	**- 3 credits Fall Semester, Senior Year**
MMSC 442 Biotechnology Practicum II:	**- 3 credits Fall Semester, Senior Year**
MMSC 443 Biotechnology Practicum III:	**- 3 credits Spring Semester, Senior Year**
MMSC 444 Biotechnology Practicum IV:	**- 3 credits Spring Semester, Senior Year**

(Non-thesis MS in Applied Molecular Biology & Biotechnology register for these courses in summer)

MMSC 641 Biotechnology Practicum I:	**- 2 credits Summer Semester**
MMSC 642 BiotechnologyPracticum II:	**- 2 credits Summer Semester**
MMSC 643 Biotechnology Practicum III:	**- 2 credits Summer Semester**
MMSC 644 Biotechnology Practicum IV:	**- 2 credits Summer Semester**

***Program Contact Number* CALL 302-831-2849 to report lateness, sick time or emergencies**

SECTION 2 – COURSE SYLLABUS

DESCRIPTION:
Practical internships in a variety of biotechnology laboratory settings. Students participate in all phases of laboratory functions.

PHILOSOPHY:
Integration of prior didactic and classroom laboratory education into varied clinical settings prepares students to become effective, professional biotechnologists. The attributes of a professional biotechnologist encompass more than those of diagnostic expertise. Laboratory scientists must be accountable not only for knowledge within their laboratory specialty, but for demonstrating dependable, ethical and disciplined behavior.

OBJECTIVE(S):
During the Clinical Practicums, students must be able to demonstrate competence in the various laboratory procedures. Students must also exhibit appropriate behaviors with respect to interpersonal relationships, dependability, integrity and professionalism. Students will have met the objectives of the Clinical Practicum courses by demonstrating competence in:

- conducting themselves in accordance with laboratory policies and procedures at each clinical site.
- exposure to and responsibility for professional behavior of a practicing biotechnologist.
- exposure to and supervised work responsibility in the biotechnology laboratory, including adjunct technologies where available and appropriate.
- accountability for accurate data analysis and documentation.
- participation in staff review of laboratory research/projects with senior scientists.
- observation of and participation in laboratory organization, including manual and/or computerized record keeping and reporting systems, quality control and quality assurance procedures and documentation methods, and personal interactions.

Overview of Core Technical Objectives

Competent use of liquid handling devices
Preparation of solutions
Use of pH meter
Preparation of media and plates
Sterilization methods
Use of spectrophotometer
Bacterial culture on both liquid and plate media
Archiving strains
Preparation of plasmid/chromosomal DNA
Analysis of recombinant plasmids
Quantitation of nucleic acid
Preparation of nucleic acids for agarose gel electrophoresis
Agarose gel electrophoresis of DNA/RNA
Restriction endonuclease digestion
Labeling of nucleic acids
Evaluation of data obtained from agarose gel electrophoresis of DNA/RNA
Proper handling/storage of nucleic acids
Transformation/Transfection
Hybridization/Blotting procedures: southern blot, western blot
Evaluation of data following blotting procedures
Immunodetection procedures
PCR
Spectrophotometric determination of protein concentrations
Polyacylamide gel preparation
Preparation of proteins for SDS-PAGE
SDS-PAGE
Detection of proteins following SDS-PAGE
Evaluation of protein data following SDS-PAGE
Quantitate cells in culture (microscopic, particle counting, flow-cytometry)
Sub-culture of cells
Recombinant protein expression
Ability to follow a written laboratory protocol
Imaging/photo-documentation of data
Proper record keeping in the laboratory notebook
Effective time management

******By the completion of all four practica, students are required to complete the technical objectives as outlined in the "Technical Objectives Checklist". FAILURE TO COMPLETE THE REQUIRED OBJECTIVES MAY DELAY COMPLETION OF THE PROGRAM.**
Students must document completion of the various objectives by date and by preceptor signature (e-signature). It is the responsibility of the student to tally completion of their objectives, and to notify the Program Director far enough in advance if it appears that additional clinical time needs to be scheduled in order to complete the objectives.****

COURSE REQUIREMENTS:

Students are required to achieve and maintain pre-determined levels of competence for technical proficiency, professionalism and correlation of theoretical and practical learning during their course of study, including the clinical practicum. Criteria and further explanation of these components can be found in specific sections of this Handbook.

Grades for the Clinical Practicums are based on:

1. **Technical performance and professionalism, as assessed by Clinical Faculty**

1. Evaluation of Technical and Professional Performance:

Professional behavior and non-diagnostic technical performance are evaluated using an evaluation instrument designed to reflect §II.B. Description of the Program, of the *Standards and Guidelines for Biotechnology Programs*. The *Standards* outline the competencies students are expected to achieve on completion of their biotechnology program. This evaluation is broken down into three parts: **(1)** affective behavior while at the rotation site (rated on a scale of 1 to 5), **(2)** ability to demonstrate basic theoretical and practical knowledge in the various areas of biotechnology (rated on a scale of 1 to 5) and **(3)** technical ability in performing various molecular biology/biotechnology procedures (rated on percent competency).

2. Completion of the Technical and Professional Skills Checklist:

Students are required to complete a variety of technical and professional activities while on practical rotation and these must be documented in the **Technical and Professional Skills Checklist. Successful completion of these activities needs to be initialed by the practicum supervisor at each site. In addition, items under categories XIII and IX require additional accompanying documentation as described below. Failure to provide documentation will result in the student not receiving credit for the activity. Documentation should be provided in the block in which the activity occurred.**

A. DEVELOP AND OFFER A BIOTECHNOLOGY EDUCATIONAL ACTIVITY

Acceptable documentation

- Copy of syllabus and/or course material, program or letter of appreciation that demonstrates content and length of teaching time Note: You can only receive credit for teaching the same topic once.

B. PAPERS, PUBLICATIONS, BOOKS, PRESENTATIONS AND EXHIBITS (PAPER or POSTER SESSIONS) INCLUDING:

- Publishing a paper in a recognized (indexed) journal or presented before a professional audience;
- Developing a technical scientific exhibit for display at a national or regional scientific meeting.

Acceptable documentation

- Title page of a publication
- Chapter listing and title page
- Abstract identifying poster session
- Meeting outline identifying presentation

C. PROFESSIONAL LEADERSHIP ACTIVITY

Participate in a recruitment/information workshop, or open house (on or off campus) promoting the Department of Medical and Molecular Sciences professions.

Acceptable documentation

- Copy of materials developed for workshop (poster/power point presentation)
- Letter of appreciation that demonstrates location, date and length of presentation.

D. ATTENDANCE AT A SEMINAR, JOURNAL CLUB, IN-SERVICE WORKSHOP, OR REGIONAL SCIENTIFIC MEETING

Acceptable documentation

- Letter or certificate of attendance or signed roster
- Title and brief description of activity with director's signature
-

COURSE GRADING:

A high level of technical proficiency is essential to Biotechnology practice. It is essential that Biotechnologists (and therefore Biotechnology students) strive to achieve the highest level of technical performance using current technology and knowledge. The expected level of

professional behavior is correspondingly high, to reflect the importance of integrity, judgment and skill required in the day to day performance expectations in the Biotech sector.

The course grading scale ***for undergraduate as well as graduate students***, is:

GRADES	Numeric Range	Quality Points
A	93-100	4.0
A-	90-92	3.7
B+	87-89	3.3
B	83-86	3.0
B-	80-82	2.7
C+	77-79	2.3
C	73-76	2.0
C-	70-72	1.7
D+	67-69	1.3
D	63-66	1.0
D-	60-62	0.7
F	Below 60	0.0
WF	Withdrew Failing	0.0

The minimum passing grade for individual practicum courses is a **C- for undergraduate students; B- for graduate students**. Undergraduate students are required to maintain a GPA of at least 2.00; graduate students are required to maintain a GPA of at least 3.00.

Grades for Practicum courses are not rounded.

SECTION 3 – DEPARTMENTAL POLICIES

DEPARTMENT POLICIES APPLICABLE TO CLINICAL/RESEARCH LABORATORY PRACTICE

Definitions:

Unsafe conduct: action(s) which poses a potential threat to the well-being, health or safety of patients, faculty, health care workers, fellow students, or self.

Unprofessional conduct: malicious, intentional or negligent action(s) which fall below, compromise or disregard the practice and ethical standards of the professional discipline, the health care or research community, and/or the educational climate.

Unsatisfactory performance: knowledge, skill(s) and/or time-in-practice insufficient to meet the minimum competencies, objectives, performance criteria, or scheduled experiences of the clinical practicum.

The determination of unsatisfactory performance, unprofessional conduct or unsafe conduct will be made by the faculty, who will determine when or if a student will be removed from or return to clinical or laboratory practice, the condition(s) for doing so, and the level of clinic or laboratory activity permitted. Depending on the severity of the incident(s) and/or number of prior incidents, the faculty's sanctions may result in dismissal from the program and/or department; repeating the clinical course; mandatory clinical time extensions; and/or remedial instruction prior to readmission to the department or re-entry into clinical or laboratory courses.

Department recommendations for dismissals based on clinical performance are subject to review and approval by the Committee on Student Promotions. Students who wish to appeal a Departmental action, including a Departmental or Program dismissal, may do so by following the provisions of the Grade Appeal Protocol.

POLICY FOR UNPROFESSIONAL OR UNSAFE CLINICAL/RESEARCH LABORATORY CONDUCT

To successfully complete each practicum course, students are expected to demonstrate clinical and laboratory competencies consistent with the policies and standard procedures taught in program courses and described in course syllabi, the College's Catalog and *Student Handbook*, and the Practicum Handbook. If, in the judgment of a clinical and/or program faculty member, the student demonstrates behavior that is detrimental to the well-being of patients, fellow students, faculty members or him/herself, the student's clinical laboratory activities will be terminated immediately. Examples of such unprofessional or unsafe conduct include, but are not limited to:

(1) tampering with, destruction or theft of equipment, specimens or teaching materials;
(2) verbally abusive, physically threatening or harmful behavior;
(3) falsification of documentation (laboratory or student records);
(4) gross interference with the educational process or health care services;
(5) gross impairment (physical or cognitive) by illicit or prescription drugs;
(6) inappropriate or unauthorized use of laboratory equipment, supplies, reagents, data, laboratory information systems, or communications systems;
(7) unsupervised clinical practice or unauthorized presence in a laboratory facility;
(8) creating unnecessary risk of exposure to or harm from environmental, chemical- and/or bio-hazards; and
(9) unauthorized, unreported and/or excessive absence during scheduled clinic time.
(10) non-compliance with the work rules, policies and/or procedures of the laboratory and/or institution.
(11) non-compliance with HIPAA, CLIA, FDA or other mandated regulatory programs, as applicable to students.

POLICY FOR UNSATISFACTORY CLINICAL PERFORMANCE

The minimum passing grade for practicum courses is C- (B- for graduate). Students demonstrating unsatisfactory clinical performance will earn a grade less than C- (B- for graduate). The letter grades of I (Incomplete) or IP (In progress) will not be used to extend an otherwise unsatisfactory rotation or practicum course.

A student who demonstrates unsatisfactory performance in a clinical practicum course must repeat that clinical course. The student will earn a grade of C- (B- for graduate) if he/she passes the repeated practicum course, or a grade of F if he/she does not pass. The repeat grade will be used to compute the grade point average. Students may repeat **only one** practicum course in this manner.

Scheduling of the repeat rotation or clinical course is subject to availability of an appropriate clinical affiliate site and adequate clinical supervision. It may be necessary for the student to wait until a rotation site becomes available. Unsatisfactory performance in the repeated rotation or clinical course may result in dismissal from the major.

EFFECT OF POLICIES ON PROGRAM COMPLETION

Students must recognize that penalties for unsafe, unprofessional and unsatisfactory performance; course failure; repeated courses; dismissals; make-up time; or additional assignments are likely to delay scheduled completion of program requirements, and may jeopardize scheduled eligibility for graduation, registry certification, and/or subsequent employment.

SECTION 4 – STUDENT AND CLINICAL SITE RESPONSIBILITIES

STUDENT RESPONSIBILITIES

1. SCHEDULING AND ASSIGNMENT OF PRACTICUM ROTATIONS

Practicum rotations are scheduled to assure (1) a broad variety of Practicum environments; (2) adequate supervision, staff interaction and representative caseload; (3) a reasonable expectation that students are able to travel to their assigned sites; and (4) that to the extent possible, student site preferences are considered during scheduling. Students may be offered the opportunity to make a preliminary selection of preferred rotation sites. In most cases, students are assigned to sites for which they have indicated a preference. However, student pre-selection of preferred rotation sites does not guarantee assignment to those sites. If the number of available Practicum sites will not accommodate all students, one or more students may be assigned to an on-site, program faculty-supervised rotation in the Department's Simulation Laboratory. Scheduling for all Practicum courses, including assignment to specific sites or times, is contingent on availability of an appropriate Practicum affiliate site and adequate supervision.

Practicum rotations (days, times and sites) are scheduled and confirmed by the Program Faculty in consultation with Clinical Faculty. No further schedule changes can be made unless (a) the student is able to demonstrate that attendance at an assigned rotation site has or will create undue or unreasonable hardship, or (b) the Clinical Instructor must alter the schedule. **In no event is the student permitted to make his or her own arrangements for** Practicum **rotations or to change scheduled rotation days, times or sites without a prior request to and approval by the Program Faculty and Clinical Faculty**.

Students are advised that even when a Practicum hardship is demonstrated, it may not be possible to assign the student to an alternate site. When this is the case, the student may choose to postpone a rotation until a site becomes available. Postponement may result in delay of program completion.

IF YOU HAVE A DISABILITY AND REQUIRE ACCOMMODATION, you must submit a request and documentation to the Office of Student Affairs. Refer to page 28 of the School of Health Professions Student Handbook.

2. TRANSPORTATION, ACCOMMODATIONS AND CLINICAL EXPENSES

Students are responsible for arranging their transportation to and from clinical sites. With few exceptions, Newark/Wilmington/Philadelphia area sites are accessible using public transportation (train, bus or subway). The Department does not have the capacity to provide students with rental cars, shuttle service, fares, tokens, or parking fees, or other cash payments for meals or accommodations at clinical sites. Students selecting or assigned to distant clinical sites must arrange their own transportation and housing.

3. HEALTH CLEARANCE

No student will be approved to begin clinical practice until he/she has demonstrated that all appropriate health requirements have been met. Requirements include documentation, physical examination, and immunizations required by the University, and any specific requirements related to Biotechnology program accreditation or professional standards. A student attending a practicum rotation without the appropriate Health Clearance will be immediately removed from the practicum site, and will not be allowed to resume his/her rotation until the Health Clearance is produced.

4. PRACTICUM ROTATION DRESS CODES

A clean, white full-length lab coat is required for all students while on rotation at University of Delaware and at most other practicum sites. Professional attire should be worn at all times during practicum rotations. **Tennis shoes, sandals, very high heeled shoes, long dresses, T-shirts, shorts and jeans are prohibited.** University of Delaware student identification badges must be worn on lab coat breast pocket. Students may wear surgical scrubs when working in clinical diagnostic labs. NOTE: Attire at practicum sites may also require lab whites and/or appropriate sterile attire to conform with CDC Universal Precautions and/or OSHA regulations for protection against transmittal of bloodborne pathogens. Students are to confirm dress codes before beginning each rotation.

5. ATTENDANCE AT ASSIGNED PRACTICUM ROTATION SITE(S)

Unless specified in the practicum **schedule, there is no "time off" from** practicums**. Students are expected to be at the rotation site during the dates and daily times scheduled. Students are required to spend a minimum of 7 hours per day of rotation, excluding breaks, lunchtime, etc. Should the student need to leave earlier than the regularly scheduled time, he or she must make arrangements to make up the time lost (ie by coming in earlier that day or other mechanism determined by the clinical instructor. Absences are recognized only for sick time, for doctor appointments that cannot reasonably be made during non-clinic hours, or for special circumstances *only when pre-approved by the Clinical Instructor and Program Faculty*. Students must inform both the BAMBB Program Office and the Clinical Faculty member at the rotation site in the event of an absence no later than 9:00 a.m. for each day of absence.**

a. Any absentee time, *including time taken for job interviews*, **in excess of eight hours over the entire clinical experience**, must be made up during the term in which the absence occurs and before a grade is recorded, unless Program Faculty expressly waive this requirement and the documentation of the waiver is in writing in the student's program file.

b. Scheduled time off **must** receive prior approval from the Program Faculty.

c. Whenever possible, absentee time should be made up at the site from which the student was absent and should be arranged with the Clinical Instructor at that site.

d. Occasionally, a Clinical Instructor will tell a student not to report to the Practicum Site on a scheduled practicum day, or will let a student leave early or come in late. **Under no**

circumstances are students to construe this as time off. When this occurs, students are to report to the Department Simulation Laboratory for that clinical day/time.

e. Program Faculty will assume absences have not been made up unless make-up time is clearly indicated on the student's worksheets, noted with the Clinical Instructor's signature.

f. Each day or part thereof of unauthorized absence will result in a 5% reduction in the final course percentage grade for the technical/professional evaluation. Students should be aware that this 5% reduction may affect successful completion of the clinical course.

6. PROFESSIONALISM

Students are expected to abide by the guidelines incorporated in their professional Codes of Ethics, and by standards and regulations applicable to clinical/research laboratory practice. Students should strive to establish good working relationships with all personnel with whom they come in contact during the Practicums. Students must demonstrate responsibility in the care of equipment and materials they use and the integrity and confidentiality of specimens they process during the assigned practicum rotations. Students should seek consultation with the Clinical Faculty member at the rotation site for problems that may arise during the practicum. In the event that a problem arises that is not resolved to the satisfaction of the Clinical Faculty member or the student, consultation will take place with the student, Clinical Faculty member and the Biotechnology Program Faculty.

7. DEPARTMENT, LABORATORY and AFFILIATE INSTITUTION POLICIES

Except as indicated in paragraph 5.d., above, students are expected to abide by the established daily work routine and attendance schedule at the Practicum rotation site or to the schedule prepared by the Program in conjunction with Clinical Faculty. If preparation or monitoring of techniques/experiments necessarily extends attendance beyond scheduled hours, it is the student's professional duty to follow through to complete the necessary work. However, **at no time is unsupervised practice or unauthorized presence in a laboratory facility permitted**. Since the purpose of practicum rotations is to maximize student exposure to and competence in laboratory practice, **the use of** practicum **time to work on other course or program assignments (e.g. research papers, class projects) is not permitted. Likewise, use of practicum site laboratory computers (for email/internet searches/text messaging), laboratory phones, or Xeroxing machines for personal reasons is not permitted. MMSC policy regarding use of cell phone and pagers remains in effect, i.e. they are not to be used while on duty – this means turn off completely.**

Student practicum performance (technical/professional components), is evaluated on a par with a laboratory position description for an entry level staff biotechnologist. Therefore, it is in the students' best interest to familiarize themselves with laboratory policies regarding employee conduct, disciplinary procedures and laboratory technical standards. Students should familiarize themselves with these policies on arrival at the rotation site.

8. DAILY WORKSHEETS: MAINTENANCE AND DOCUMENTATION

Maintenance of work records and accurate documentation of work product are essential to practice in biotechnology laboratories. The Biotechnology Program provides blank daily worksheets to students and to Clinical Instructors. Each student is responsible for maintaining a **LABORATORY NOTEBOOK**, in which **ALL LABORATORY ACTIVITY MUST BE ENTERED AND DOCUMENTED for each day of rotation**. ***The student should determine if the laboratory prefers the student to use his/her own laboratory notebook or whether the student should supply his/her own.*** IN ANY CASE **ALL STUDENTS MUST HAVE A LABORATORY NOTEBOOK**. **The laboratory notebook, as it contains IP of the laboratory in which you work becomes the property of the lab.** In addition, the student must complete a daily log which is to be signed off by your immediate supervisor on a weekly basis. To satisfactorily document actvities, **LABORATORY NOTEBOOK** AND the electronic **DAILY LOG** Worksheet must include and clearly indicate the date, and the nature of the work carried out on a given day. **It is the student's responsibility to submit to the Program Director his/her electronic assessments for review and evaluation no less than seven (7) calendar days after completion of each practicum course and/or as required for Program review**. Please see the "Forms" section for the log form for paper representations of the assessments. Specific instructions to students for completing electronic worksheets will be provided in the practicum orientation meeting.

Students must return these electronic forms no more than seven (7) calendar days after completion of each rotation. ***Failure to accurately document practicum work or to submit worksheets in a timely manner may result in significant point deductions, delay of grade reports or failure of the Clinical Practicum course*****.**

REQUIRED ELECTRONIC ASSESSMENTS TO BE INCLUDED IN BOUND FORM PACKET

- **Professional and Technical Evaluation form signed by preceptor.**
- **Daily logs initialed by supervisor.**
- **Technical skills competency checklist.**
- **Completed professional/technical activities log sheet.**
- **Student Evaluation of Site.**
- **Practicum Absence log sheet (as applicable).**

9. CLINICAL AFFILIATE SITE ASSESSMENT

Students evaluate rotation sites as part of our reciprocal evaluation procedure. Students must return these forms to the Program office no more than seven (7) calendar days after completion of each rotation. Please see the "Forms" section for the form.

Anonymous, composite evaluations, completed by students are returned to each site at the completion of rotations for each academic year. A copy is maintained in the Program's Practicum Site files.

10. EMPLOYMENT INTERVIEWS

Students should make every effort to schedule appointments for job interviews on days when practicums and classes are not scheduled. However, students **in good standing may** be approved for a maximum of one practicum day (8 hours) for a job interview(s) **only** if the following conditions are understood and met. Note that the eight hour maximum spans the entire practicum phase of the program. ***This policy should not be construed to mean one day off within each clinical course.***

a. A request for interview time off must be submitted to the Program Faculty at least one week in advance of the tentative date of the interview.

b. Program Faculty must pre-approve requested time off for interviews.

c. Sick leave and/or required clinical time can not be used or substituted for interview time.

d. **Time off granted for interviews in excess of eight (8) hours must be made up**. Time approved for interviews during regularly scheduled classes or clinical rotations does not excuse students from meeting requirements for that class or clinical rotation, including required time in clinical practice.

e. Program Faculty determine where and when missed time for job interviews will be made up.

11. CAREER DEVELOPMENT CENTER

The University's Career Development Center offers a variety of career-related services, free of charge, to students of the Dept. of MMSC. The Center will help you set short and long range career goals, prepare a resume, write letters (such as cover and thank you letters), make contacts and schedule employment interviews, prepare for interviews, evaluate job offers, select a graduate program, and investigate financing for graduate education.

- The Career Development Center keeps a list of job opportunities available to UD students and graduates, including part-time work for students and full time professional positions for graduates of each program.
- The Center also provides the computerized career planning program Discover, which guides you step by step through the career evaluation and planning process.
- The Career Development Center has evening hours by appointment.

12. WEATHER EMERGENCY POLICY

Should weather conditions necessitate, the University of Delaware may declare a weather emergency.

Once a weather emergency is declared, all on-campus and off-campus classes (clinical and non-clinical) are cancelled.

- Students scheduled to be at off-campus clinical locations should contact their immediate clinical supervisor at the rotation site to inform him/her of the UD Weather Emergency.
- UD Weather Emergencies are announced on local radio stations* and posted on the UD website. ***Call 302-831-8109 for Department-specific information*.**

13. STUDENT PROFESSIONAL LIABILITY COVERAGE

The University of Delaware maintains insurance coverage for professional and general liability for all matriculated students while they are on authorized practicum affiliate assignments. Only students officially registered for clinical courses are covered by this policy. Only when participating in activities specifically designed for the practicum or other approved courses are students covered by this policy.

CLINICAL AFFILIATE SITE & CLINICAL/PROGRAM FACULTY RESPONSIBILITIES

Biotechnology Clinical Faculty at clinical affiliate sites share responsibility with Program Faculty and the students themselves for the professional education of students enrolled in the Department of Medical and Molecular Sciences. The Clinical Faculty occupy a key role in making the students' practicum experience a successful and meaningful one.

Practicum sites maintain active affiliate status by providing at least one student rotation experience in each academic term (i.e.: during each of the Fall, Spring, Winter-session, Summer semesters). The list of active clinical affiliate sites is updated annually. All biotechnologists employed at active sites are eligible to attend Department of Medical and Molecular Sciences-sponsored continuing education workshops, conferences, seminars and other activities for substantially reduced or no fees. Biotechnologists employed at inactive clinical affiliate sites may attend Department-sponsored activities at the regular fee.

Clinical Faculty work closely with the Program Faculty and are responsible for:

1. serving as a model of the professional for students to emulate.
2. orienting students to the corporate and/or laboratory facilities, and to the personnel, policies, and procedures involved in the day to day functioning of their laboratory.
3. insuring that students read the policy and procedure manual and abide by the employee conduct guidelines and laboratory standards therein.
4. supervision, technical and diagnostic instruction, and evaluation of students during student rotations at the Practicum site with respect to work assigned to and completed by the student.

5. reviewing, verifying and initialing student *Daily Worksheets* on a regular basis during the rotation and at the completion of the rotation prior to making a final assessment of the student's performance.
6. clearly and accurately indicating on student worksheets and on the *Evaluation Forms* the basis for awarding or deducting points for student technical and professional performance. ***Note: Clinical and Program Faculty are required to comply with the evaluation methods and assessment standards as outline in this Handbook.***
7. providing a signed evaluation of the student's competency attainment and professional performance based on the guidelines provided by the Program or based on an evaluation system established by the practicum site in conjunction with Program Faculty.
8. assuring that the *Technical and Professional Evaluation* of each student reflects a factual and objective assessment of the student's cognitive, motor and affective abilities and behaviors. [Students are evaluated on a par with an entry level Staff Tech position]
9. conferring with Program Faculty throughout the academic year at regular intervals regarding students' performance, and review of students' individual worksheets.
10. attending Clinical Affiliate meetings to assure currency with evaluation and accreditation requirements
11. submitting as appropriate updated annual laboratory statistics and personnel data to the Program for the Program's required institutional and/or accreditation reports.

SECTION 5 – STUDENT FORMS

Note: Students are required to complete electronic versions of these forms – hard copies of these are provided for student review purposes only.

College of Health Sciences
Department of Medical and Molecular Sciences
Program in Biotechnology/Applied Molecular Technologies

Practicum Supporting Materials Checklist

Name:_____ Date:_____

Practicum Course Number:_____ Rotation Site/Preceptor:_____

Item	**Disposition**
Preceptor Evaluation of Student	OYes ONo (if no, when_____)
Student Daily log with signatures	OYes ONo (if no, when_____)
Technical Competency Checklist	OYes ONo (if no, when_____)
Student Evaluation of Site	OYes ONo (if no, when_____)

I understand that failure to complete/submit the requirements will not allow for assignment of a grade for the given practicum.

Signature of Student_____ Date: _____

Signature of Program Director (or designate)_____Date:_____

College of Health Sciences
Department of Medical and Molecular Sciences
PROGRAM IN BIOTECHNOLOGY/APPLIED MOLECULAR TECHNOLOGIES

STUDENT DAILY ACTIVITY LOG

Student _____ _____ **Site:** _____

AVOID POINT DEDUCTIONS: CHECK HANDBOOK TO MAKE SURE DATA IS ENTERED CORRECTLY

PRACTICUM ROTATION TIME VS. RESEARCH PROJECT TIME MUST BE CLEARLY INDICATED

PROFESSIONAL ACTIVITY TIME MUST BE NOTED HERE AND ON P/T ACTIVITIES LOGSHEET

	MONDAY	TUESDAY	WEDNESDAY	THURSDAY	FRIDAY
	Date_____	Date_____	Date_____	Date_____	Date_____
8-9A					
9-10A					
10-11A					
11-12A					
12N-1P					
1-2P					
2-3P					
3-4P					
4-5P					
5-6P					

STUDENT DAILY ACTIVITY LOG

I have reviewed the above named student's daily log and laboratory note book and believe it to be an accurate representation of the student's activities in the lab.

SUPERVISOR SIGNATURE:_____ **DATE:**_____

COLLEGE OF HEALTH SCIENCES
DEPARTMENT OF MEDICAL AND MOLECULAR SCIENCES DATE DUE _____
Program in Applied Molecular Biology and Biotechnology

TECHNICAL & PROFESSIONAL EVALUATION
TO BE COMPLETED BY PRACTICUM INSTRUCTOR

Student_____ Clinical Site_____

Rotation Dates: From____/___/___to____/___/___Practicum Instructor _____

Instructions to Evaluator: The columns indicate numerical grades and equivalent letter grades. Please OBJECTIVELY indicate, by assigning a numerical grade within one column, the level of competence at which this student performed in each category while on rotation in your laboratory. (Eg: 86% would be entered under column D) This checklist is a COMPREHENSIVE LIST – NOT ANY ONE LABORATORY WILL BE PERFORMING ALL OF THE LISTED TASKS. If you feel a category or sub-category is not applicable to your laboratory's situation, please mark "N/A".

Graduate /Undergraduate: **A A- B+ B B- C+ C C- D+ D D- F**

I. GENERAL LABORATORY SKILLS					
A. Demonstrated ability to properly use pH meter: selected proper buffers, performed appropriate calibration techniques, took proper care of electrodes, cleaned/maintained pH meter					
B. Demonstrated ability to use spectrophotometer: selected proper cuvette, wavelength, and parameters for calibration; allowed ample time for bulbs to warm up and displayed proper shut-down/clean up procedures					
C-1. Used autoclaving sterilization methods successfully: chose proper glassware/containers for autoclaving, used autoclave sensitive tape, ran machine according to type of sample being autoclaved					
C-2. Used filter sterilization methods successfully: chose proper filter type, decontaminated area properly, displayed proper use of biological safety cabinet, practiced aseptic/sterile technique					
D. Prepared all reagents/solutions accurately: ensured proper labeling, followed aseptic and sterilization procedures when appropriate, stored correctly					
E. Demonstrated proper use of liquid handling/measuring devices (pipettors, pipets, glassware, etc): measured and dispensed volumes accurately, disposed of pipets properly, cleaned/disinfected as needed					
F. Performed all tasks as per laboratory protocol: understood technical vocabulary, selected and appropriately used equipment, supplies, reagents, and samples; recalled previous demonstrations to perform work independently; used proper controls; disposed of waste correctly					
G. Displayed ability for competent record keeping in either laboratory notebook or other paperwork: entries were clear, concise, complete and accurate; writing was legible, no "white-out" used					
H. Captured images via camera or other photodocumentation equipment: produced images that were clear and well focused, followed safety procedures for UV light exposure					
II. MICROBIOLOGY					
A. Prepared liquid/plate media effectively: calculated correct concentrations, displayed proper pH and autoclaving techniques, practiced aseptic/sterile technique					

Graduate /Undergraduate:

	A+	A	A-	B+	B	B-	C+	C	C-	D+	D	D-	F
B. Performed bacterial culture of both liquid and plate media: ensured accurate pH, used correct media, selected appropriate specimens													
C. Archived strains: used correct container and solutions, stored at appropriate temperature, labeled tubes with needed information													
D. Prepared plasmid/chromosomal DNA: demonstrated ability to properly follow procedure for extraction													
E. Analyzed recombinant plasmids: extracted DNA successfully, digested DNA with proper restriction enzyme, evaluated DNA on gel													
III. CELL CULTURE													
A. Performed transfection/transformation: used correct DNA concentration, exposed sample to appropriate temperature for correct time, grew transformants correctly													
B. Subcultured cells: practiced aseptic/sterile techniques, used biological safety cabinet, selected proper specimens for splitting, labeled each specimen as per protocol, provided routine care in a timely manner													
C. Performed recombinant protein expression: prepared culture with correct antibiotic use, inoculated media, corrected growth to correct OD, evaluated using SDS-PAGE/Western blot													
D. Quantitated cell in culture (microscopic, particle counting, flow cytometry): selected appropriate methods and parameters/ reference ranges; established equipment alignment and calibration; demonstrated proper safety procedures for waste, biohazards, laser hazards; evaluated data properly													
IV. HYBRIDIZATION													
A. Performed hybridization/blotting/microarray/ELISA, IFM procedures: used correct methodology; selected proper reagents and supplies; demonstrated proper electrophoresis safety													
B. Evaluated blotting/microarray/ELISA, IFM data: determined successful versus failed procedures/determined need for repeat experiments; correctly interpreted data													
C. Performed immunodetection procedures: selected correct antibodies and reagents/supplies; used correct methodology													
V. PROTEIN BASED METHODS													
A. Calculated protein concentrations using spectrophotometer: displayed proper use and maintenance of equipment; performed calculations/analyzed data accurately													
B. Prepared proteins for SDS-PAGE: exposed samples to proper temperature, selected proper dyes and markers													
C. Performed SDS-PAGE electrophoresis: assembled apparatus correctly and selected proper voltage; practiced electrophoresis safety; dissembled/cleaned apparatus correctly; disposed of gel appropriately													
D. Evaluated SDS-PAGE gels: used proper staining/destaining techniques; incubated for correct time; used light box appropriately, determined successful and failed experiments, performed photodocumentation													
VI. NUCLEIC ACIDS													
A. Extracted nucleic acids: evaluated sample acceptability, performed extraction successfully, labeled sample completely, stored appropriately													
B. Quantitated nucleic acids: selected proper cuvette type and wavelength, performed calculations correctly													

Graduate /Undergraduate: **A+ A A- B+ B B- C+ C C- D+ D D- F**

C. Prepared nucleic acids for agarose gel electrophoresis: selected appropriate dye type and amount for each sample, selected correct molecular weight marker						
D. Electrophoresed DNA/RNA on agarose gel: make gel correctly as per protocol, assembled/dissembled/cleaned apparatus properly, practiced electrophoresis safety						
E. Evaluated data from agarose gel electrophoresis: determined need for repeat/failed procedures, evaluated gel correctly, differentiated between "true" bands, versus background, primer dimers, etc.						
F. Performed PCR: determined proper reagent amounts for mix, selected correct primers and instrument settings, worked at correct temperatures, guarded against cross contamination, stored PCR products at correct temperature						
G. Digested DNA with restriction endonuclease: selected proper enzyme and calculated proper dilution of enzyme, added appropriate amount of enzyme for PCR product and incubated at correct temperature for adequate time						
H. Prepared polyacrylamide gel: made gel correctly, assembled/dissembled/cleaned apparatus, practiced electrophoresis safety						
I. Handled/stored nucleic acids properly: demonstrated proper procedure to avoid contamination, stored sample at proper temperature.						
J. Set up and run RNA based analysis such as Real-Time or Digital PCR reaction.						
VII. BIOINFORMATICS						
A. Performed literature database search: chose appropriate search engines, selected key words to yield best results, retrieved applicable articles						
B. Performed search for gene/protein/DNA information: demonstrated ability to successfully navigate web, chose appropriate sites, participated in on-line tutorials, modeling demonstrations/exercises						
VIII. BEHAVIOR						
A. Followed all local, state, and federal regulations concerning the handling, storage, and disposal of chemicals and biohazardous materials						
B. Operated equipment properly and safely, maintained cleanliness of equipment and workspace, recorded preventive maintenance, temperature logs, and other performance records						
C. Practiced discretion, scientific accountability, and confidentiality with laboratory and patient records.						
D. Demonstrated honesty and integrity in daily duties, and truthfulness in relationships with peers and staff, showed interest in what they were doing						
E. Practiced good interpersonal communication skills with peers, faculty, and laboratory personnel						
F. Accepted constructive criticism, modified behavior accordingly in response to supervision, followed directions carefully, showed maturity in dealing with problems						
G. Demonstrated dependability in and accountability for the clinical experience and work environment, including scheduled attendance, punctuality, adherence to daily work schedules, prior notice for absences, assuring missed times was made up according to program requirements						
H. Adhered to all personal protective equipment (PPE) regulations, including wearing gloves, laboratory coat, and other protection as needed; changed when soiled; disposed of appropriately.						
I. Concentrated on work, did not use computers for personal use, avoided excessive use of phone for personal calls, did not use rotation time to study/do class work.						
J. Demonstrated ability to multi-task; showed initiative to find work during "down-time"; expressed interest in laboratory activities						

Graduate /Undergraduate: A+ A A- B+ B B- C+ C C- D+ D D- F

MISCELLANEOUS LABORATORY-SPECIFIC ACTIVITIES (NOT LISTED ELSEWHERE)					

STUDENT AVERAGE (BASED ON SCORES ABOVE) _____

CORRESPONDING LETTER GRADE _____

Using a checkmark [ü]**, please rate this student in the following areas <u>in comparison to</u>**
(A) other University of Delaware students, and
(B) other students as indicated below
(*please check all that apply*).
_____ **Undergraduates (other than University of Delaware)**
_____ **First/second year MS or PhD UD graduate students**
_____ **Terminal year MS or PhD non-UD graduate students**
_____ **Medical students/residents**

	Below Average (Lowest 40%)	**Average** (Middle 20%)	**Above Average** (Next 20%)	**Good** (Top 20%)	**Outstanding** (Top 10%)	**Truly Exceptional** (Top 5%)	**Inadequate opportunity to Observe**	**A UD Students** / **B Other Students**
Technical Ability								A
								B
Theoretical Knowledge								A
								B
Ability to apply knowledge/skills to appropriate procedure								A
								B
Communication skills: Oral								A
								B
Communication skills: Written								A
								B
Ability to analyze problems and formulate solutions								A
								B
Maturity								A
								B
Motivation / Perseverance								A
								B

1. Were there circumstances that may have influenced your evaluation of this student? Explain.

2. Were there circumstances that may have adversely influenced the student's performance? Explain.

3. At this time, how would you rate this student for employment in your area on an overall evaluation?

[ ] Highly recommended (98-100%) **[ ] Not recommended (less than 90%)**

[ ] Recommended (90-97%) **[ ] Unable to evaluate**

Practicum Instructor Signature _____

Date _____

Has this evaluation been reviewed with the student? [ ] YES [ ] NO

UNIVERSITY OF DELAWARE
COLLEGE OF HEALTH SCIENCES
DEPARTMENT OF MEDICAL AND MOLECULAR SCIENCES
Program in Applied Molecular Biology and Biotechnology

TECHNICAL AND PROFESSONAL SKILL SETS

TO BE COMPLETED BY STUDENT

Student _____ Practicum Site(s):

Block I Rotation dates _____ to _____ _____

Block II Rotation dates _____ to _____ _____

Block III Rotation dates _____ to _____ _____

Block IV Rotation dates _____ to _____ _____

Instructions to Student: The columns indicate activities that you performed at each of your rotation sites. Each skill has a number enclosed in parentheses. This number is the MINIMUM number of that activity that should be completed at the end of your combined rotations. When an individual activity is performed, a tick mark should be placed in the corresponding rotation box. Tick marks should be placed only to represent **SUCCESSFUL completions – do not count unsuccessful attempts**. At the end of each rotation, your clinical instructor should initial each box where ticks are present, to confirm this accurately reflects work performed at that site. There are some activities for which there is overlap between two different skills listed. When this is the case, please check only one skill and make a notation on the other skill set (*eg*. When recording work performed determining protein concentration using a spectrophotometer, MARK THE ACTIVITY IN **PROTEIN BASED METHODS – A,** and then note in **GENERAL LABORATORY SKILLS – B** to "SEE PROTEIN BASED METHODS A.")
All rotation information should be recorded on the same checklist – do not use a different checklist for each block! At the end of the final rotation, you should add up activities from all rotations and place in the "total" box, to ensure that minimum amount was met for all activities. **MINIMUMS SERVE AS GUIDELINES ONLY – IF YOU HAVE COMPLETED THE'MINIMUM', YOU SHOULD NOT TAKE THIS AS NOT HAVING TO DO ANY MORE!!!**

All sub-tasks for each skill should be completed for a "Success"	**Block: I**	**II**	**III**	**IV**	**Total**
I. GENERAL LABORATORY SKILLS					
A. USE OF PH METER (4) 1. Select proper buffer for calibration 2. Calibrate correctly 3. Use magnetic stirrer for solution 4. Take proper care of electrodes 5. Display correct shut-down					
B. USE OF SPECTROPHOTOMETER (4) 1. Select proper cuvette for specimen type 2. Program proper wavelength/other parameters 3. Allow ample warm-up time for bulbs 4. Calibrate/standardize as appropriate using proper controls 5. Clean machine after use.					
C-1. USE OF STERILIZATION METHODS – AUTOCLAVING (4) 1. Select proper glassware/containers 2. Label with autoclave tape 3. Run accordingly to type of material being autoclaved					
C-2. USE OF STERILIZATION METHODS-FILTER STERILIZATION (4) 1. Select proper filter type 2. Display proper use of biological safety cabinet 3. Decontaminate area 4. Practice sterile/aseptic technique					

All sub-tasks for each skill should be completed for a "Success"	Block: I	II	III	IV	Total
D. PREPARATION OF SOLUTIONS (12) 1. Use scales/volumetric glassware accurately 2. Use magnetic stirrer while mixing 3. Allow sufficient time for ingredients to dissolve completely 4. pH as appropriate 5. QC/QA as needed 6. Label with name of solution, date, expiration date,etc. 7. Store solutions properly					
E. COMPETENT USE OF LIQUID HANDLING DEVICES (PIPETORS, SEROLOGICAL PIPETS, GRADUATED CYLINDERS, ETC) (20) 1. Measure volume accurately (proper meniscus placement, etc) 2. Dispense volume completely 3. Dispose of pipets and other disposables properly 4. Clean/disinfect pipettors as needed					
F. ABILITY TO FOLLOW WRITTEN LABORATORY PROTOCOLS (20) 1. Understand technical vocabulary 2. Select appropriate reagents, supplies, and equipment 3. Remember basic steps of protocol when demonstrated, in order to repeat with minimal supervision 4. Select proper controls 5. Dispose of waste correctly/demonstrate proper clean-up					
G. PROPER RECORD KEEPING IN A LABORATORY NOTEBOOK/ OTHER LAB SPECIFIC PAPERWORK (20) **1. Record information in pen, no "white-out" use, corrections are crossed out, dated and initialed** **2.** Data is complete, informative, and concise **3.** Data is legible					
H. IMAGING/PHOTODOCUMENTATION OF DATA (20) 1. Select proper camera settings (exposure time, aperture opening, etc) 2. Allow sufficient development time 3. Ensure picture is well focused, producing clear pictures with crisp images 4. Follow safety method for UV light exposure and gel handling					
I. ATTEND LABORATORY MEETINGS/SEMINARS/ CONTINUING EDUCATION ACTIVITIES (8)					
II. MICROBIOLOGY					
A. PREPARATION OF LIQUID/PLATE MEDIA (4) 1. Calculate concentration of media 2. pH media 3. Autoclave media 4. Practice sterile/aseptic technique					
B. BACTERIAL CULTURE OF BOTH LIQUID AND PLATE MEDIA (4) 1. Ensure pH is accurate 2. Select appropriate media 3. Select appropriate cells					
C. ARCHIVING STRAINS (4) 1. Choose container for storage 2. Select proper solutions for preservation 3. Store at appropriate temperature 4. Label with expiration date, if applicable					
D. PREPARATION OF PLASMID/CHROMOSOMAL DNA (8) 1. Follow proper procedure (commercial kit or organic {phenol}) for extraction 2. Evaluate DNA on agarose gel					
E. ANALYSIS RECOMBINANT PLASMIDS (8) 1. Extract DNA successfully 2. Digest DNA with proper restriction enzyme 3. Evaluate DNA fragments on agarose gel					

All sub-tasks for each skill should be completed for a "Success"	Block: I	II	III	IV	Total
III. CELL CULTURE					
A. TRANSFORMATION/TRANSFECTION (4) 1. Use correct concentration of DNA 2. Expose sample to appropriate temperature/time for heat shock 3. Grow transformants properly					
B. SUBCULTURE OF CELLS (4) 1. Practice sterile/aseptic technique 2. Select proper cultures ready for sub culturing 3. Ensure proper labeling of each subculture 4. Provide routine care of cultures in a timely manner					
C. RECOMBINANT PROTEIN EXPRESSION (4) 1. Prepare overnight culture 2. Inoculate media 3. Demonstrate correct antibiotic use 4. Correct growth to correct OD 5. Use inducing agent (ex. IPTG) 6. Evaluate using SDS-PAGE or Western blot					
D. CYTOMETRIC TECHNIQUES (8) (quantitative cells in culture {microscopic, particle counting, flow cytometry}) 1. Select appropriate method for analysis 2. Establish reference range/parameter criteria 3. Perform quality control, including software manipulation alignment, calibration, and preventive maintenance 4. Demonstrate proper safety procedures for waste, biohazardous materials, laser hazards. 5. Evaluate data correctly					
IV. HYBRIDIZATION					
A. HYBRIDIZATION/BLOTTING PROCEDURES, WESTERN BLOT, MICROARRAY, ELISA, IFM (4) 1. Use correct methodology 2. Select proper reagents, antibodies, blocking and washing solutions 3. Use proper electrophoresis safety methods					
B. DATA EVALUATION FOLLOWING BLOTTING PROCEDURES (4) 1. Determine failed procedures/need for repeat experiment 2. Analyze and accurately evaluate results					
C. IMMUNODETECTION PROCEDURES (4) 1. Select proper antibodies SEE HYBRIDIZATION – A FOR ADDITIONAL INSTRUCTIONS					
V. PROTEIN-BASED METHODS					
A. SPECTROPHOTOMETRIC DETERMINATION OF PROTEIN CONCENTRATIONS (4) 1. Select proper cuvette type and instrument wavelength/ parameters 2. Calibrate/standardize instrument 3. Analyze data accurately 4. Demonstrate proper clean-up/shut-down of instrument					
B. PREPARATION OF PROTEINS FOR SDS-PAGE (12) 1. Expose sample to appropriate temperature treatment 2. Select appropriate dyes and markers to use					
C. SDS-PAGE ELECTROPHORESIS (12) 1. Assemble apparatus correctly 2. Select proper voltage 3. Practice electrophoresis safety 4. Disassemble/clean apparatus correctly 5. Dispose of gel appropriately					

All sub-tasks for each skill should be completed for a "Success"	Block: I	II	III	IV	Total
D. EVALUATION OF SDS-PAGE (12) 1. Use of proper staining/destaining procedures 2. Incubate for appropriate time 3. Use of light box 4. Evaluate need for repeat/failed samples 5. Photodocumentation					
VI. NUCLEIC ACIDS					
A. NUCLEIC ACID EXTRACTION (10) 1. Evaluate specimen type and volume acceptability 2. Perform extraction by manual or kit oriented methods 3. Ensure end product tube is accurately labeled with appropriate information 4. Store appropriately					
B. QUANTITATION NUCLEIC ACIDS (4) 1. Select proper cuvette type and instrument wavelength parameters 2. Perform calculations correctly ALSO SEE GENERAL LAB SKILLS-B AND PROTEIN BASED METHODS-A					
C. PREPARATION OF NUCLEIC ACIDS FOR AGAROSE GEL ELECTROPHORESIS (12) 1. Select appropriate dye type and amount 2. Select appropriate molecular weight marker					
D. AGAROSE GEL ELECTROPHORESIS OF DNA/RNA (12) 1. Make determination of gel type (low melting, high melting) and concentration and make gel accordingly 2. Allow gel to completely polymerize 3. Assemble apparatus and run at proper voltage 4. Practice electrophoresis safety 5. Allow adequate time for running completely 6. Disassemble/clean apparatus appropriately 7. Dispose of gel correctly					
E. EVALUATION OF DATA OBTAINED FROM AGAROSE GEL ELECTROPHORESIS OF DNA/RNA (12) 1. Determine failed/need to repeat procedures 2. Evaluate data correctly 3. Differentiate between "true" bands, versus background, primer dimers, contamination					
F. REAL-TIME or DIGITAL PCR (2) Wet bench or approved virtual+assessment 1. Set up of reaction 2. Program thermocycler/operate software 3. Analyze data to determine technical proficiency.					
G. PCR (12) 1. Determine proper reagent amounts 2. Select correct primers 3. Work at correct temperatures (on ice after addition of Taq, etc) 4. Program instrument correctly 5. Take appropriate measures to guard against sample mix-up and cross contamination 6. Store PCR products appropriately					
H. RESTRICTION ENDONUCLEASE DIGESTION (12) 1. Select proper enzyme and make working enzyme dilution 2. Add appropriate amount of working enzyme to DNA/PCR product 3. Incubate at correct temperature for specific enzyme					

I. POLYACRYLAMIDE GEL PREPARATION (3) 1. Select concentration and make gel accordingly 2. Assemble apparatus correctly 3. Allow sufficient time for gel to polymerize 4. Run gel at proper voltage 5. Practice electrophoresis safety 6. Dissemble/clean apparatus appropriately					
	Block:				
All sub-tasks for each skill should be completed for a "Success"	**I**	**II**	**III**	**IV**	**Total**
J. PROPER HANDLING/STORAGE OF NUCLEIC ACIDS (8) 1. Demonstrate proper procedures to prevent contamination of sample 2. Ensure tube is properly labeled 3. Store sample based on elution buffer to prevent DNA degradation (eluted in H2O must be frozen, eluted in Tris-HCl or other kit buffers can be stored short term in refrigerator)					
VII. BIOINFORMATICS					
A.PERFORM LITERATURE DATABASE SEARCH (2) 1. Choose appropriate search engines 2. Select key-words that will yield best search results 3. Retrieve applicable articles and print hard copies as needed					
VIII. PROFESSIONAL DEVELOPMENT ACTIVITIES (8)* **1. Attendance at a seminar** **2. Attendance at a Journal Club** **3. Attendance at an In-Service Workshop** **4. Attendance at a regional scientific meeting** **5. Presentation at a regional scientific meeting** **6. Publication of a paper in a indexed journal** **7. Development and presentation of a technical demonstration for display at a regional or scientific meeting.**					
IX. LEADERSHIP DEVELOPMENT ACTIVITIES (4)* **1. Participation in a recruitment/information workshop, or open house (on or off campus) promoting Bioscience Technologies professions.** **2. Develop and offer a Biotechnology educational activity**					
X. MISCELLANEOUS LABORATORY-SPECIFIC ACTIVITES (NOT LISTED ELSEWHERE)					
***Note – Activities listed under sections VIII and IX must have accompanying documentation. See section 2 for guidance. Any combination of activities under sections VIII or IX is acceptable.**					

UNIVERSITY OF DELAWARE
COLLEGE OF HEALTH SCIENCES
DEPARTMENT OF MEDICAL AND MOLECULAR SCIENCES
Program in Applied Molecular Biology and Biotechnology

PRACTICUM ABSENCE LOG SHEET

Name Student (print):_____

Name Student (signature):_____

DATE ABSENT	PRACTIUM NUMBER	HOURS/DAYS ABSENT	DATE/HOURS MADE UP

UNIVERSITY OF DELAWARE
COLLEGE OF HEALTH SCIENCES
DEPARTMENT OF MEDICAL AND MOLECULAR SCIENCES
Program in Applied Molecular Biology and Biotechnology

PRACTICUM ORIENTATION MEETING
ACKNOWLEGEMENT AND ATTENDENCE DOCUMENT

I have attended the Program in Applied Molecular Biology and Biotechnology Practicum Orientation Meeting and **understand** the professional, ethical, attendance and course requirements (including submission of documentation forms) for the practica.

I understand that failure to comply with these policies may adversely affect my grade(s) for the clinical practica and/or completion of the practica course sequence.

Name Student (print):_____

Name Student (signature):_____

Date:_____

Faculty:_____

Date:_____

UNIVERSITY OF DELAWARE
COLLEGE OF HEALTH SCIENCES
DEPARTMENT OF MEDICAL AND MOLECULAR SCIENCES

DATE SENT _____

DATE DUE _____

Program in Applied Molecular Biology and Biotechnology

CLINICAL PRACTICUM SITE EVALUATION - TO BE COMPLETED BY STUDENT

Practicum Site _____ Practicum Instructor _____

Student: _____ Rotation dates_____to _____

Rotation Block (circle one): I II III IV

Instructions to Student: The columns indicate categories in which you should assess your experience at this particular site. Please evaluate each category twice: (A) as a practicum site, and (B) as it compares to your previous rotation sites. If this is your first rotation, please mark "N/A" in the comparison (B) rows.

	Below Average	Average	Above Average	Truly Exceptional	Inadequate Opportunity to Observe	A This site Evaluation / B compared to your other sites
1. Site was prepared for my arrival and my practicum experience.						A
						B
2. Professional behavior was demonstrated in the laboratory.						A
						B
3. Adequate supervision was provided; personnel explained procedures as needed; were available for questions						A
						B
4. Communication between lab personnel and me was beneficial and appropriate						A
						B
5. Lab resources (patients/ specimens/equipment/space, etc) were adequate.						A
						B
6. My experiences at this site met Practicum course objectives.						A
						B
7. Site provided activities for me to minimize "down-time"; involved me in daily activities.						A
						B
8. Site was clean, free of clutter, and adhered at all applicable regulations (Proper disposal of waste, PPE, etc.)						A
						B
9. Activities contributed to my knowledge and development of technical skills.						A
						B

Provide additional comments about this site, including explanation for "below average" ratings.

Describe this site's strengths:

Describe this site's weaknesses/areas for suggested improvement:

Would you recommend this site to others? Why or why not?

Please use other side for additional comments

UNIVERSITY OF DELAWARE
School of Health Professions / Department of Medical and Molecular Sciences
Program in Applied Molecular Biology and Biotechnology
STUDENT EVALUATION OF PRACTICUM

Check Applicable - - Practicum I - - Practicum II - - Practicum III - - Practicum IV

Rotation: - - Other *(Specify)*_____

Student Name: [please print] _____

SITE_____TERM_____YEAR_____

Name(s) of those to whom you were directly responsible *[list first and last name(s)]*:

_____

I. Overview: Check the description that most closely represents your evaluation of the facility.

1. Were your student responsibilities and privileges reviewed with you?
   - _____ Very clearly presented.
   - _____ Adequately discussed. Knew what was expected of me on a day-to-day basis.
   - _____ In general, I knew what was expected of me on a day-to-day basis. Occasionally unclear as to my responsibilities.
   - _____ Unclear. Left confusion in my mind as to what was expected of me.

2. Were you conscious of a well-planned program for students in this facility?
   - _____ Excellent program. The site kept students occupied with pertinent work, allowing student to feel productive.
   - _____ Good program. Student usually well occupied with pertinent work.
   - _____ Adequately planned program. Student assigned pertinent tasks but work flow was somewhat slow.
   - _____ Poorly planned program. Student was not assigned sufficient tasks to keep occupied. Student experienced a more than average amount of slack time.

3. Do you feel that the responsibilities you were given at this facility were adjusted to your ability to handle them?
   - _____ The responsibilities given me were suited to my ability to handle them. They were appropriate for a newly graduated biotechnologist entering the workforce.
   - _____ Some of the responsibilities were above my ability to handle them. In my opinion, they were appropriate for a more experienced technologist.
   - _____ I felt competent in handling all responsibilities given me. However, in general, the amount of responsibility given me was somewhat limited and therefore not appropriately adjusted to my abilities.
   - _____ The responsibilities given me were not adjusted to my ability. The responsibilities were too limited and the amount of experience too narrow.

4. Do you feel the facility allowed flexibility for the individual student to gain maximum benefits?
   - _____ The student program was quite flexible. Students were encouraged to pursue additional tasks or interests when routine or assigned work was completed.
   - _____ The student program was somewhat flexible. If the student demonstrated a desire to pursue additional tasks or interest once assigned work was completed, the laboratory staff was helpful.
   - _____ Due to the amount and nature of the work the student did not have the opportunity to pursue additional tasks.
   - _____ A strict protocol was followed each day. Once assigned work was completed students were sent home.

II. Supervision and Instruction

Please rate the facility on each item below by circling the appropriate number on the rating scale.

The rating scale is:

0	1	2	3	4	5
not applicable	poor	adequate	above average	very good	outstanding

A. Apparent interest in student progress	**0**	**1**	**2**	**3**	**4**	**5**
B. Supervision of student	**0**	**1**	**2**	**3**	**4**	**5**
C. Fostering of student learning	**0**	**1**	**2**	**3**	**4**	**5**
D. Amount of feedback given student	**0**	**1**	**2**	**3**	**4**	**5**
E. Receptivity toward students' question	**0**	**1**	**2**	**3**	**4**	**5**

III. Practicum Experience

1. List below the instruments and other major equipment you operated.

_____

_____

_____

_____

2. List the types of procedures you observed but did not perform.

_____

_____

_____

_____

3. What additions and or deletions would you make to the program at this facility? Please explain.

_____

_____

_____

IV. Academic/Practicum Correlation

1. Did you find correlation between previously learned theories and concepts and their practical application at this facility? If your answer is "no", please explain.
( ) yes ( ) no

_____

_____

_____

2. What specific recommendations would you make to more successfully correlate your learning experience within the University with the practical experience in this facility?

_____

_____

_____

V. Student's Signature _____

Dates of Practicum _____

Date of Evaluation _____

UNIVERSITY OF DELAWARE
COLLEGE OF HEALTH SCIENCES
DEPARTMENT OF MEDICAL AND MOLECULAR SCIENCES

DATE SENT _____

DATE DUE _____

Program in Applied Molecular Biology and Biotechnology

STUDENT PRACTICUM PERFORMANCE SELF-EVALUATION

Practicum Site _____ Practicum Instructor _____

Student: _____ Rotation dates_____to _____

Rotation Block (circle one): I II III IV

Instructions: **Use this form to evaluate your own performance at this site.** Columns indicate numerical grades. Indicate, by assigning a **numerical grade within one column**, the level of competence at which you believe you performed in each category while on rotation in this laboratory. (eg: 86% would be entered under column labeled "89 87 85")

Graduate /Undergraduate:	A+ 100	A 99	A- 98	B+ 97	B 96	B- 95	C+ 94	C 92	C- 90	D+ 89	D 87	D- 85	F <85
A. I followed all local, state, and federal regulations concerning the handling, storage, and disposal of chemicals and biohazard materials.													
B. I operated equipment properly and safely, and maintained cleanliness of equipment and my workspace.													
C. I practiced discretion and confidentiality with lab and patient records in accordance with HIPAA requirements and professional ethics.													
D. I was honest and truthful in relationships with peers and staff. I demonstrated integrity in my daily duties. I showed interest in the lab's policies and procedures and understood the lab's workflow.													
E. I practiced good interpersonal communication skills with peers, faculty, and laboratory personnel.													
F. I accepted constructive criticism, modified my behavior accordingly in response to supervision and followed directions carefully. I showed maturity in dealing with personal or lab problems.													
G. I was dependable and accepted responsibility for my practicum experience and work environment, including scheduled attendance, punctuality, adherence to daily work schedules, prior notice for absences and assuring missed time was made up. I was present at this site at least 7 hours each day, exclusive of lunch and breaks.													
H. I was knowledgeable about and adhered to the policies, procedures and instructions as indicated in my Practicum Handbook													
I. I adhered to all personal protective equipment (PPE) regulations, including wearing gloves, laboratory coat, and/or other protection as needed when present in prep/wet laboratories or special procedures.													
J. I concentrated on my work. I did not use lab computers for personal use. I did not make or receive personal calls during lab time. I did not use rotation time to work on non-clinical class assignments.													
K. I demonstrated ability to multi-task; showed initiative to find work during "down-times"; expressed interest in laboratory activities.													
L. I learned and practiced applying principles of quality control and quality assurance as required by current laws and regulations.													

Additional comments about your performance at this site: